#### Medicinal values of the genus Phyllanthus - A review

Unigwe, Cyprian Robinson<sup>1\*</sup>, Ukwueze, Chigozie Stanley<sup>2</sup>, Okey, Stephen Nnaemeka<sup>1</sup>

<sup>1</sup>Department of Veterinary Biochemistry and Animal Production, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria.

<sup>2</sup>Department of Veterinary Medicine, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria.

Submitted: 15<sup>th</sup> July., 2023; Accepted: 19<sup>th</sup> July., 2023; Published online: 31<sup>st</sup> Aug., 2023 DOI: https://doi.org/10.54117/jcbr.v3i4.5 \*Corresponding Author: Unigwe Cyprian Robinson; <u>robinsonunigwe@gmail.com</u>, +2348037707965

## Abstract

Phyllanthus species are known for their many medicinal values used in folklore medicine in many parts of the world. They have array of phytochemicals in the leaf, flower, seed, stem and root, that confer medicinal advantage on them. Extracts have shown a broad range of pharmacological activities like hepatoprotective, antioxidant, antiviral, antimicrobial, antidiabetic, anti-inflammatory, anticancer, antimalarial, nephroprotective, diuretic, and several other properties. This review highlighted different research results from other scientists in the past and present, all tailored towards exploring the inherent pharmacological properties of this herb for ethno-medicine, alternative medicine and pharmaceutical purposes.

**Keywords**: Folklore medicine, Herb, Pharmacological activity, *Phyllanthus* species, Therapeutic use **Running title:** Medicinal values of *Phyllanthus* 

## Introduction

*Phyllanthus* is a native genus largely used as antipyretic, antiviral, antidiarrheal, antispasmodic, antinociceptive, antiinflammatory, antioxidant, antitumoral, antigenotoxic, antimutagenic, hepatoprotection, hypoglycemic, hypotensive, diuretic, antidiabetic and bactericidal agent (Maria et al., 2019). It was also efficient for jaundice, hepadnaviral fighting DNA polymerase, AIDS and kidney disorders (Mao et al., 2016; Devi et al., 2017; Meena et al., 2018). Phyllanthus amarus Schum and Thorn and Phyllanthus niruri L. have been frequent targets of ethnopharmacological work all over the world, mainly in Brazil, U.S.A, Malaysia, Cuba, Peru, Caribbean, China, Nigeria, Africa and India (Devi et al., 2017; Meena et al., 2018).

*Phyllanthus* sp. contains bioactive compounds, flavonoids, including terpenes, lignins. phenols, saponins, terpenoids, alkaloids, and tannins (Bahar et al., 2011; Hidanah et al., 2022). This plant has a natural antioxidant, antimicrobial, and immunomodulator, which will increase the immune system's components and improve the immune system's function (Hidanah et al., 2018). Bioactive compounds have antimicrobial (Adeolu and Sunday, 2013; Unigwe et al., 2021), anti-inflammatory (Dada et al., 2014; Unigwe et al., 2020), anti-diabetic (Evi and Degbeku, 2011), antioxidant (Lim and Murtijaya, 2007), anticonvulsant activity (Manikkoth et al., 2011), anti-diarrhoeal

(Unigwe *et al.*, 2021), anti-carcinogenic and anticancer (Rajeshkumar *et al.*, 2002) as well as antiviral (Salazar *et al.*, 2011; Prasad *et al.*, 2013).

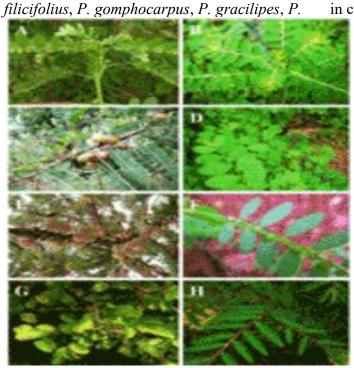
The most dominant bioactive compounds in Phyllanthus sp. are hypophyllanthin, catechin, epicatechin, rutin, and quercetin, and chlorogenic, ellagic, caffeic acid, malic, and gallic acids (Mediani et al., 2017). These compounds have been reported to have antibacterial properties (Mujeeb et al., 2014). Phyllanthus niruri L. extract inhibited the growth of Escherichia coli, which was indicated by a clear zone at 19.3 mm (Lestariningsih et al., 2015a). P. niruri L. powder at a level of 1.2% can minimize E. coli in the broiler intestine (Lestariningsih et al., 2015b). It was also reported that 65% P. niruri L. extract at a dose of 1 mL/kg body weight, administered orally for 7 days reduced white blood cells in chickens infected with Mycoplasma gallisepticum (Hidanah et al., 2018). The administration of 10 g/kg Phyllanthus amarus powder to chickens not infected with bacteria decreased the number of leucocytes but increased the red blood cell count (Unigwe et al., 2020). When 100 mg/mL concentrations of the water, 50% ethanol and petroleum ether extracts of P. screened for preliminary amarus was mycobactericidal activity, the water and 50% ethanol extracts showed activity against Mycobacterium smegmatis with a zone of inhibition of 1.5  $\pm$  0.2 and 2 cm  $\pm$  0.5 respectively. P. amarus had a minimum inhibitory concentration (MIC) of 50 mg/mL and 100 mg/mL for the 50% ethanol and aqueous extracts respectively, while rifampin had an MIC of 0.1 µg/mL (Bekoe et al., 2020). Therefore. this review was aimed at underscoring the medicinal values hidden in Phyllanthus species of herb, which is easily seen around our surrounding as weed.

#### History of *Phyllanthus amarus*

Phyllanthus niruri also known as Phyllanthus amarus Schum and Thonn is a member of Euphobiaceae family. The origin of Phyllanthus amarus is tropical America; from there it spread as a weed to other tropic and sub-tropics. It is a tropical annual herb shrub which grows as weed in moist humid waste land (Adeneye and Senebo, 2008). P. niruri is among more than 500 Phyllanthus species that are widely spread in temperate and tropical climates region (Iizuka et al., 2007). It grows 30 - 40 cm in height, has small leaves and yellow flowers; the stem has green capsule, and blooms with flowers with 5 white sepals and apical acute anther. The fruit has green capsules, and smooth and fruiting pedicels while seeds are longitudinally rugose (Obianime and Uche, 2009). It is found throughout the tropics and sub- tropics such as West Africa (including Nigeria and Ghana), Europe, Asia (including China, Pakistan, India and Malaysia indian ocean), central and south America as medicinal plant for the treatment of various diseases (Etta, 2008; Jain et al., 2008). The plant has been used for a long period of time (thousands of years) in Ayurvedic traditional medicine for various illnesses (Adedapo et al., 2005). In India, Phyllanthus niruri is one of the most important traditional medicines used for the treatment of jaundice, asthma, hepatitis and urolithic disease (Ishimaru et al., 1992).

the Phyllanthus is largest genus of Phyllanthaceae family. It is further subdivided into 11 subgenera including Gompidium, Bortryanthus, Conani, Cicea, Emblica, Ericocus. Isocladus. Kirganelia, Phyllanthodendron, Xyllophylla, and 1,270 Phyllanthus. species There are of Phyllanthus distributed throughout most of the subtropical and tropical regions including

tropical Africa, tropical America, Oceania, and Asia (Hoffmann et al., 2006). About Phyllanthus species 200 are widely distributed in tropical America, mainly in the Caribbean and Brazil (Unander et al., 1995). There are 20 species of Phyllanthus commonly found throughout Malaysia which include P. amarus, P. albidiscus, P. chamaepeuce, P. columnaris, P. elegans, P. emblica, debilis. Ρ. *P*.



oxyphyllus, P. pachyphyllus, P. pulcher, P. reticulates, P. ridleyanus, P. roseus, P. sikkimensis, P. urinaria, P. virgatus, and P. watsonii (Burkill, 1966). Of all these, Phyllanthus species, P. amarus, P. emblica, P. urinaria, P. niruri, Phyllanthus acidus L., P. fraternus, P. reticulatus, and P. simplex are widely used in traditional medicine to heal immune-related diseases and their reported immune-modulating properties in cell, animal, or human studies.

**Figure 1:** *Phyllanthus* species (Phyllanthaceae) pictorials; (A) *P. amarus*, (B) *P. urinaria*, (C) *P. emblica*, (D) *P. niruri*, (E) *P. acidus*, (F) *P. fraternus*, (G) *P. reticulatus*, and (H) *P. simplex*.

*P. amarus* Schum and Thonn (synonym: *P. niruri* Auct) is found in tropical and subtropical regions. It is an erect annual herb, usually found in fields, grasslands, and forests and is erect annual herbs, 10–60 cm tall. Leaves distichous, flowers small in leaf axils, 1–2 together. Perianth segments 5 or 6, green with broad scarious margins, enlarged in fruiting. Stamens 3; filaments entirely connete. Styles bifid at the apex. Fruits globose, trigonous, depressed at the apex. Seeds 3-gonous, with 5–7 sub-parallel longitudinal ribs (Mitra and Jain, 1985).

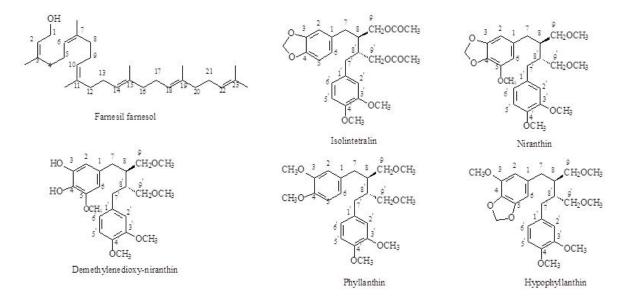


Figure 2: Chemical structures of some Phyllanthus amarus bioactive constituents.

#### Source: Maria *et al.* (2019)

Table 1: Phytoche	nicals in Phyllanthus amarus
-------------------	------------------------------

Compound	Constituent
Alkaloids	Isobubbialine and Epibubbialine (Houghton et al., 1996)
Tannins	Geraniin, corilagin, 1,6-digalloylglucopyranoside rutin, quercetin-3-O-
	glucopyranoside, Amarulone, Phyllanthusiin D and Amariin (Foo and Wong,
	1992; Foo, 1993).
Lignans	Niranthin, Nirtetralin, Phyltetralin, Hypophyllanthin, Phyllanthin,
	demethylenedioxy-niranthin, 5-demethoxy-niranthin, Isolintetralin (Kassuya et
	al., 2006; Leite et al., 2006; Maciel et al., 2007; Srivastava et al., 2008).
Ellagitannins	Amariin, 1-galloyl-2,3-dehydrohexahydroxydiphenyl (DHHDP)-glucose,
	Repandusinic acid, Geraniin, Corilagin, Phyllanthusiin D, and flavonoids namely
	rutin, and quercetin 3-O-glucoside, 1 Ogalloyl- 2,4-
	dehydrohexahydroxydiphenoyl-glucopyranose elaeocarpusin, repandusinic acid
	A and geraniinic acid (Foo, 1995; Londhe et al., 2009)
Volatile oil	Linalool and Phytol (Moronkola et al., 2009).
Tritepene	(2Z, 6Z, 10Z, 14E 18E, 22E-farnesil farnesol) (Maciel et al., 2007).

P. niruri Linn. (synonyms: Diasperus chlorophaeus (Baill). Kuntze. Nymphanthus niruri (L). Lour., P. carolinianus Blanco, P. chlorophaeus Baill., P. parvifolius Steud., P. purpurascens Kunth., P. rosellus (Müll.Arg). Müll.Arg., *P. williamsii* Standl). It is a small erect annual herb growing up to 30–40 cm in height and is indigenous to the Amazon rainforest and other tropical areas, including South East Asia,

JCBR Vol 3 Is 4 July-Aug 2023

Southern India, and China. Its leaves are 7–12 cm long, and they are alternate, sessile oblong. It has small off-white-greenish flowers, which are solitary, auxiliary, pedicellate, apetalous, and monoecious (Mitra and Jain, 1985). P. niruri is closely related to P. amarus in appearance and phytochemical contents, but a recent cladistic analysis indicated that the Phyllanthus genus is paraphyletic, and therefore, these two problematic and confusing species, P. niruri and P. amarus, are two individual species (Lee et al., 2006).

Р. urinaria L. (synonym: Р. leprocarpus Wight) is widely distributed in pantropic areas. It is a monoecious and shortlived perennial herb and may grow up to 60 cm tall. Erect, annual, glabrous herbs, leafbearing branchlets short, flattened, or slightly winged. Leaves are distichous, closely placed. Flowers are minute, axillary, solitary, subsessile. Stamens are 3; filaments connect into a column. Fruits depressed globose, echinulate and scarcely lobed. Seeds are trigonous, rounded on back, transversly rugose (Kuttan and Harikumar, 2011).

Р. emblica L. (synonym: Emblica officinalis Gaertner). Distributed throughout tropical and subtropical India, wild or planted. It is a deciduous tree. The tree is small to medium in size, reaching 1-8 m in height. Young branch lets reddish brown, hairy, leaves apiculture obtuse, rounded, or chordate at base, glabrous beneath, about  $18 \times 1.2$  cm. Flowers are greenish vellow, usually monoecious. Fruit fleshy more than 1.5 cm in diameter is nearly spherical, light greenishyellow, quite smooth, and hard on appearance, with six vertical stripes or furrows. The branchlets are not glabrous or finely pubescent, 10-20 cm long, usually deciduous; the leaves are simple, subsessile and closely set along branchlets, light green, resembling pinnate leaves (Kuttan and Harikumar, 2011).

P. acidus L. (synonyms: P. distichous L., Cicca acida L., Cicca disticha L). is native to Malaya Islands and Madagascar and is found throughout Asia and also in the Caribbean region, Central and South America. It is a deciduous trees reaching 2 to 9 m high. Leaves are distichous, light green above, glaucous beneath. Flowers are densely clustered and elongated, simple or branched racemes from old, defoliated branches, male flowers numerous, female 1-2, stamens 4, much longer than perianth. Ovary is 3-4 locular. Fruits fleshy, depressed globose, 6-8 lobed, pale yellow. Fruits borne in loose clusters are pale yellow or white, waxy, crisp, juicy, and very sour with edible small yellow (Kuttan and Harikumar, 2011).

*P. fraternus* G.L. Webster (synonyms: *P. fraternus* subsp. *togoensis* Jean F. Brunel & J.P. Roux *P. lonphali* Wall.) distributed in Tropical Asia especially in Indian subcontinent and Pakistan. It is an erect, annual plant growing up to 45 cm, occasionally to 60 cm (Balakrishnan and Chakrabarty, 2007).

P. reticulatus Poir (synonym: Kirganelia reticulata Poir). An Asian species of *Phyllanthus* distributed throughout tropical India. It is a large, glabrous or pubescent, straggling shrub which grows up to 5 m high. Flowers are auxiliary; the male flowers in fascicles of 2-6, the females are solitary. Perianth segments 5-6, oblong, alternatively with disk glands. Stamens are 5, the inner 3 with their filaments connete into column, the outer 2 free, shorter. Ovary is 5-10 locular, styles 3, minute. Fruits fleshy, depressed globose, dark purple or black, shining. Seeds 8-10, trigonous granulate (Kawakita and Kato, 2009).

*P. simplex* Retz. (synonyms: *P. virgatus* G. Forst). Distributed in South Asia; India, Sri Lanka to SE. Asia, S. China, Indo-China, and

Malesia to Pacific Islands. It is fairly common along roadsides, in open fields and dry deciduous forests. Erect or diffuse perennial herb, up to 20 cm high. Leaves are distichous, subsessile. Flowers are usually solitary, on slender axillary pedicels. Perianth segments oblong, obtuse, slightly enlarged in fruit. Stamens are 3, distinct; anthers didymous. Style short, bifid. Fruits are globose, 3-lobed. Seeds trigonous, rounded on back, covered with minute tubercles in irregular lines, dark brown (Kuttan and Harikumar, 2011).

#### **Ethnomedicinal values**

Several Phyllanthus species have been utilized for a long time by traditional healers of India, China, Brazil, and countries in the Southeast Asian regions potent as ethnomedicines to treat numerous diseases including those related to the human immune system. P. amarus, P. emblica, P. niruri, P. reticulatus, Phyllanthus corcovadensis, and P. fraternus are among others that are widely used as ethnomedicines in many parts of the world. In Brazil and in many South American countries, the infusion of roots, stems, and leaves of most *Phyllanthus* species have been used to cure a broad spectrum of diseases including intestinal infections, hepatitis B, diabetes, kidney, and urinary bladder disturbances (Calixto et al., 1998). In Asia, several Phyllanthus species are used as febrifuge, diuretic, deobstruent, stomachic, and antiseptic. Ayurveda uses the most number of Phyllanthus species where 15 species have been used in the management of genitourinary, hypertension, cancer, skin, digestive, hepatic, and respiratory disorders (Nain et al., 2012; Sarin et al., 2014). Among these, P. niruri is the mostly used as a remedy for inflammation, fever, malaria, lithiasis, gonorrhea, and hepatitis. P. amarus is used widely in Ayurvedic medicine for the treatment of stomach, liver, kidney, genitourinary system and spleen problems,

menorrhagia, gonorrhea, and other genital infections. It is also beneficial in intermittent fevers. diarrhea. ulcers and wounds. gastropathy, dysentery, ophthalmopathy, and scabies (Bagalkotkar et al., 2006; Ifeoma et 2013). In Thailand. al.. six *Phyllanthus* species are used extensively as ethnomedicines. P. urinaria, P. amarus, and P. virgatus are used for the treatment of gonorrhea, diabetic, jaundice, and hepatic disorders while the other species, P. niruri, P. acidus, and P. reticulatus are used to treat hypertensive, malarial fever, constipation, and skin and urination disorders. In China, five species are widely used in Tibetan and Traditional Chinese Medicine to treat anuria, dropsy, hypertension, sore throat, and hepatitis B as well as blood and bile disorders. Particularly, P. niruri, P. simplex, and P. reticulatus are used for the treatment of inflammation, urinary infection, rheumatism, and ophthalmic diseases (Nain et al., 2012). In Africa, six Phyllanthus species are used to treat tetanus, malarial fever, and wound (Omulokoli et al., 1997; Ifeoma et al., 2013). Specially, in Cameroon, Ghana, Ivory Coast, Kenya, Nigeria, and Zambia, the stem, root, leaves of various species and of Phyllanthus are used for the remedy of tetanus, wound, fever. and sexually transmitted diseases.

## Phytochemistry

Many phytochemical, biological, and pharmacological studies of Phyllanthus species have been performed due to tremendous ethnomedicinal and therapeutic potentials of the plants. More than 510 compounds have been isolated and identified from several Phyllanthus species. Most of the species were reported to comprise of diverse combinations of secondary and bioactive metabolites including lignans, triterpenes, sterols, alkaloids, flavonoids, ellagitannins, and other polyphenols. These

compounds provide important medicinal properties to the plants. There are already a few reviews on the phytochemistry and pharmacological properties of Phyllanthus species (Calixto et al., 1998; Nahar et al., 2011; Patel et al., 2011; Sarin et al., 2014). Specially, the lignans and tannins isolated from genus Phyllanthus have been evaluated for their wide variety of biological activities. Corilagin (1), geraniin (2), and gallic acid (3) are the three most predominant metabolites found in this genus. Mostly, corilagin (1), geraniin (2), gallic acid (3), phyllanthin (4), hypophyllanthin (5), ellagic acid (6). phyltetralin (7), niranthin (8), catechin (9), quercetin (10), astragalin (11), and chebulagic acid (12) are the bioactive metabolites that have been intensively investigated and reported to exhibit diverse biological and pharmacological effects including immunomodulatory activities (Jantan et al., 2014).

## Chemical components of Phyllanthus

An initial phytochemical exploration of Phyllanthus species reported the occurance of terpenoids, alkaloids, glycosides, flavonoids, tannins, and saponins (Mao et al., 2016; Lee et al., 2016). Phenolic compounds, especially tannins, are the major constituents of Phyllanthus plants. More than 100 phenolic constituents with diverse biological activities were comprehensively identified in the fruits of P. emblica L. using HPLC-MS (Yang et al., 2012). It is also emphasized that different parts of Phyllanthus plants have different isomers of the same compounds. Phyllanthus species are rich in phytochemical diversity, with compounds tannins, such as phenylpropanoids, terpenoids, phenolic compounds, flavonoids, alkaloids, saponins and many of their glycosides. Almost 81 been compounds have isolated from Phyllanthus spp. During 2016–2018, the

majority of which were phenylpropanoids, triterpenoids, diterpenoids, and flavonoids.

# The components isolated from each *Phyllanthus* species

# a) Phenylpropanoids

Phenylpropanoids constitute the most prevalent class of compounds in the genus Phyllanthus. Thirty seven compounds were identified from P. glaucus (Nguyen et al., 2016), P. amarus (Muthusamy et al., 2016; Pereira et al., 2016 ; Pereira et al., 2017), P. urinaria (Muthusamy et al., 2016) and P. brasiliensis (Borges et al., 2018). Among these compounds lignans such as neolignan, norlignan and sesquineolignan were the most prominent. Interestingly, most of the compounds were present in the form of enantiomers in P. glaucus, including nine pairs of enantiomeric lignans 1-18.

# b) Terpenoids

Terpenoids are another major class of chemicals in the genus Phyllanthus. About 19 compounds including 11 triterpenoids 38–48), diterpenoids 49–55. one seven and monoterpene 56) were identified in P. hainanensis (Fan et al., 2017) and P. urinaria (Chung et al., 2016; Wu et al., 2017). It is noteworthy that compound 38 is a new skeleton compound, which incorporates an unprecedented 6/9/6 heterotricyclic system in the lower-left and a highly oxygenated 5,5spirocyclic ketal lactone motif in the upperright. Compounds 42–48 are lupine type pentacyclic triterpenoids.

# c) Phenolic Compounds

Seven phenolic compounds, including one new mucic acid 1-ethyl 6-methyl ester 2-Ogallate (57), together with six known phenolic compounds such as gallic acid (58), methyl gallate (59), ellagic acid (60), 1,6-di-Ogalloylglucose (61), mucic acid 1,4-lactone methyl ester 5-O-gallate (62), and mucic acid dimethyl ester 2-O-gallate (63) were isolated from the fruits of *P. emblica* L. (Zhang *et al.*, 2017, Yan *et al.*, 2017). The gallotannins corilagin (64) and geraniin (65) were also isolated from *P. niruri* and *P. muellerianus* respectively (Boakye-Gyasi *et al.*, 2016).

## d) Flavonoids

Compounds **66–69** are kaempferols, which contain one kaempferol (**66**) and three kaempferol glycosides (**67–69**), and were identified from *P. acidus* (Tram *et al.*, 2016). Quercetin (**70**) and its glycoside rutin (**71**) were found in extracts of both *P. niruri* L. (Kaur *et al.*, 2016) and *P. amarus* (Putakala *et al.*, 2017). Two new chalconoid analogues with anti-tobacco mosaic virus (TMV) activity, namely emblirol A (**72**) and B (**73**), and flavanol catechin (**74**) were isolated from the roots of *P. emblica* L. (Yan *et al.*, 2017).

## e) Alkaloids

Five securinega alkaloids including (+)-allonorsecurinine (**75**), ent-norsecurinine (**76**), nirurine (**77**), bubbialine (**78**), and epibubbialine (**79**) were isolated from *P*. *fraternus* G.L. Webster (Komlaga *et al.*, 2017).

## Biological activities of *Phyllanthus* i) Parasitology

The study of parasites and their relationships with the host, called parasitology, has been a quite vast area of research in the last few decades. A lot of studies have focused on plants to find active ingredients that can fight various pathogens and stop their activity in fraternus G.L.Webster the host. Р. (Phyllanthaceae) is enriched with various alkaloids such as the securinega alkaloid (+)allonorsecurinine and many other previously known alkaloids. These compounds showed highly antiplasmodial activity against chloroquine-resistant (W2) and -sensitive (3D7) strains of Plasmodium falciparum (Komlaga et al., 2017). Higher oxidative prominently stress features in the pathogenesis of malaria, especially anaemia and patho-physiological modifications in certain body organs. Seed extracts of P.

amarus, chloroquine (CLQ) and artesunate (ATS) may effectively reduce this oxidative stress alone or in combination with various vitamins (A, B, C and E) in Plasmodium berghei-infected mice. The highest antioxidant activity was shown by Phyllanthus seed extracts alone or in combination with vitamins (A, B, E) in P. berghei (NK 65 strain) infected mice. The combined activity of artesunate/vitamins also showed an enhanced antimalarial activity due antioxidant their activity, while to combination with vitamin С was counterproductive (Ojezele et al., 2017). Human schistosomiasis is an important but neglected disease in tropics caused by blood flukes (Schistosoma spp.), which affects around 0.3 million people annually. The only known reported treatment is the use of paraziquantel (PZQ). De Oliviera et al. (2017) tried crude hexane (HE) and ethanolic (EE) extracts of P. amarus in mice infected with Schistosoma mansoni (BH strain).

in vivo schistosomicidal The activity evaluation of mice fed with extracts of P. amarus once, for two different infection periods at 30 and 45 days post-infection showed that histopathologically, granuloma decreased in both number and size for groups treated with 250 mg/kg of HE (45 dpi) or EE (30 or 45 dpi). Both HE and EE of P. amarus have antischistosomal activities, however, they act differentially according to the parasites' age. The schistosomicidal activity results in groups treated 30 days postinfection is extremely important since praziquantel does not show any activity against the juvenile forms of Schistosoma spp. (De Oliviera et al., 2017). P. amarus HE and EE extracts showed promising results against S. mansoni in vivo (De Oliviera et al., 2017). The dried fruits of Emblica officinalis (syn. Phyllanthus *emblica*) showed antitrypanosomal activity and cytotoxic effects in vitro. Vero cell line cells were

incubated with *Trypanosoma evansi* over 12 h and treated with various concentrations (250~1000 g/mL) of *E. officinalis* for an invitro cytotoxicity assay. A sharp decrease in trypanosome number was observed at 250 g/mL concentration, and trypanosomes were completely killed after 5 h of treatment. This is statistically equivalent to the 4th hour of diminazine aceturate (Berenil) treatment, the standard reference drug used. *E. officinalis* dried fruits demonstrated a potential pathway for the development of new trypanocides if in-depth investigations were to be put in place (Peter *et al.*, 2016).

## ii) Cardiovascular protection

A large number of people across the globe suffer from heart/stroke attacks, mainly due to life styles and increased poor sugar/carbohydrate intakes. In a recent study, the cardioprotective action of aqueous extracts of P. amarus was studied against high-sugar (fructose) diet-mediated cardiac damage in Wistar rats. Following 60-days of sugar diet, heart and aorta tissue samples were collected for further histopathological and biochemical analyses. Co-administration of P. amarus aqueous extracts plus glucose-diet for a specified time period (60 days) inhibited cardiac and aortic lipids levels (total lipids, triglycerides, total cholesterol and free fatty acids) and reduced phospholipid formation (Putakala et al., 2017). Histopathological evaluations of the heart and aorta tissues highlighted that the plant aqueous extracts treatment lessened the deposition of fats and necrosis. This study showed the obvious cardioprotective potential of P. amarus aqueous extract for treatment of high sugardiet mediated oxidative stress in rats is mainly due to its ameliorative antioxidant potential with its antihyperglycemia and along antihyperlipidemic properties (Putakala et al., 2017). Moreover, the phytochemical profiling of the aqueous extracts of P. amarus indicated it may contain the flavonoids quercetin and phyllanthin rutin. the lignans and hypophyllanthin, saponins and the phenolic compound gallic acid, all of which are potent drivers of cardioprotective action (Putakala et al., 2017). In another study, a combined ethanolic extract of P. emblica fruits with an ethanolic extract of Alpinia galanga rhizomes (7:3) showed a strong synergistic antioxidant response against various reactive oxygen species in endothelial cells (ECV304) in a dose-dependent fashion, and caused an increase in cell viability. It was found that the potent and active ingredient is quercetin, which at 10 g/mL concentration reduced the H<sub>2</sub>O<sub>2</sub>-mediated lipid peroxidation. When combined ellagic acid with and hydroxycinnamic derivatives, quercetin may aid antioxidant-mediated cytoprotection (Chansriniyom et al., 2018).

## iii) Antioxidant Activity

Antioxidants are active compounds that have the potential to mitigate the oxidation and degradation of various cellular proteins and ingredients by reactive oxygen species (ROS). Antioxidants play a crucial role in good health as in at-rest conditions ROS and antioxidants maintain a steady balanced level in the body (Devi et al., 2016). Medicinally important flora have played a role in resolving health issues throughout the world for centuries, and have currently gained international focus in recent decades. The presence of antioxidant molecules in plants is well documented and there is an ever increasing demand for natural synthetic additives antioxidants over (Akporowhe et al., 2016). Normal metabolism oxvgen and exogenous of factors continuously generates free radicals i.e. ROS which may initiate a cascade of chemical reactions that could be damaging to the human body (Rusmana et al., 2017).

Once these chemical cascades are initiated *in vitro* they may produce damage that leads to a decrease in number of viable cells (Singh *et al.*, 2016). The fruit and leaf extracts of *P. acidus* have been reported to possess a variety

of antioxidants, which are well known to quench ROS generated by the cellular metabolism, and check oxidative stressmediated ailments such as cardiovascular diseases and inflammation (Angamuthu et al., 2016). Moreover, leaf water extracts of P. amarus are desirable due to their maximum yield of the biologically potent antioxidant compounds for in vitro antioxidant activity, ROS scavenging and inhibition of lipid peroxidation (Nguyen et al., 2017). Singh and colleagues studied ethanolic and aqueous extracts of P. niruri to find antioxidant activity by using DPPH (2,2-diphenyl-1picrylhydrazyl) radical and H<sub>2</sub>O<sub>2</sub> scavenging assays (Singh et al., 2016). A standard antioxidant (ascorbic acid) was used as control to compare the aqueous extract antioxidant efficacy and confirmed the strong antioxidant activity, however, ethanolic extracts of P. amarus and P. niruri showed higher ROS scavenging response compared to aqueous extracts (Singh et al., 2016; Ramandeep et al., 2017).

Many studies have confirmed the presence in P. niruri extracts of quercetin, which is considered as potential natural antioxidant which is even more powerful antioxidant than vitamin E (Dai et al., 2016; Rusmana et al., 2017). P. acidus fruit ethanol and water extracts were also studied to find antioxidant and cytotoxic levels, which showed that water extracts are more potent antioxidant and cytotoxic compared to fruit ethanolic extracts (Andrianto et al., 2017). Different solvents viz. acetonitrile, alcohols like ethanol and methanol. water. ethyl acetate. and dichloromethane were used through multiple extraction methods such as conventional, ultrasound-assisted, microwave-assisted, etc. to determine the extraction efficiency and antioxidant capacity of P. amarus (Nguyen et al., 2016). Moreover, water and microwaveassisted extraction has been demonstrated as the most efficient solvent and technique for the maximum isolation of biologically active compounds from P. amarus for wide application in industry (Nguyen et al., 2016). The aqueous extracts of three endemic Cuban Phyllanthus species, viz. P. chamaecristoides microdictyus Urb., Urb., *P*. and Р williamioides Gr. Were examined to determine their antioxidant, photoprotective and antimutagenic activities (Menéndez-Perdomo et al., 2017). This study claimed that DNA damage was reduced by Phyllanthus aqueous extracts and is not linked with desmutagenic effect in vitro. and genoprotective activity is due to the induction of expression of DNA repair proteins, reduction of ROS and related mechanisms (Menéndez-Perdomo et al., 2017). Among the various *Phyllanthus* species,  $P_{\cdot}$ chamaecristoides extracts have higher antioxidant activity than those of P. microdictyus and P. williamioides. It is generally believed that the higher antioxidant potential of any compound ultimately lowers the UVR mediated oxidative damage (Menéndez-Perdomo et al., 2017). Zhang et al. (2014) reported two new compounds (bisabolane-type sesquiterpenoid diphenyl ether derivatives) along with previously 23 known compounds from the fruit extracts of *P*. emblica. These two new isolates were tested for antioxidant capability through DPPH assays and for cytoprotective effects and showed activity against H<sub>2</sub>O<sub>2</sub>-mediated injury to PC12 cells (Zhang et al., 2014).

Long term exposure to pesticides may cause severe effects to most body organs, which have been attributed to an elevation of ROS and genomic DNA damage. P. emblica was also reported to be beneficial for in vivo protection against the effects of fungicidal pesticides such as captan, which causes generate genotoxicity and help ROS (Noorudheen and Chandrasekharan, 2016). The extracts could mitigate captan-mediated oxidative stress and genotoxicity which is presumed to be due to the potential antioxidant activity of the P. emblica extracts. P. indofischeri leaves and bark extracts were prepared in water and ethanol, and found to possess significant amylase inhibitory and antioxidant activity (Kalpana et al., 2016). The methanol leaf extracts had better activity than bark extracts, hence providing a clue that P. indofischeri can be a useful addition to use as a potent antioxidant and hypoglycemic candidate to combat ROS-mediated ailments. The methanolic leaf extracts of *P. emblica* were examined, and it was said that the polyphenolics present in it may provide strong protection against lipid peroxidation damages, and increased levels of SOD, glutathione peroxidise and catalyse enzymes are primarily due to the presence of gallic acid, rutin, caffeic acid, and kaempferol (Tahir et al., 2016; Sabir et al., 2017).

*P. muellerianus* (Kuntze) Exell., is used as a wound healing agent in different African countries.

Geraniin, a powerful antioxidant, was identified as a major constituent in the aqueous extracts of *P. muellerianus* that help reduce oxidative stress and boost the healing process in chronic wounds (Boakye et al., 2016). The effects of geraniin in treating chronic wounds are mainly due to elevation of SOD, CAT and APx levels, and decreased malondialdehyde (MDA) levels at the wound site. Moreover, geraniin significantly and efficiently reduced ferric ion in vitro, which helped block the iron-mediated amplification of ROS (Boakve et al., 2016). The ethanolic leaf extracts of P. amarus were studied to find the cause of modification in serum antioxidant levels along with ROS-mediated MDA in mice, and it was reported that the antioxidant defense capacity and revitalization of the blood in treated mice was increased, which might due to the presence of previously reported compounds (Akporowhe and Onyesom, 2016). P. urinaria has a well known history in traditional medicine systems

to treat cancers, particularly osteosarcoma, which is one of the most aggresive cancers of bones. Osteosarcomas originally arise from the primitive transformed cells of mesenchymal origin that form malignant osteoids (Huang et al., 2016). The aqueous preparations of P. urinaria affect human osteosarcoma in vivo and significantly reduced tumorigenesis, but have no effect on the rest of the body organs. The reduction in tumor development was suggested to mainly be due to mitochondrial dysfunction linked with dynamic changes brought up by apoptosis and anti-angiogenesis induced by P. urinaria (Huang et al., 2016). The in vitro anti-oxidant and anti-proliferation potential of the water extracts made from the aerial parts and roots of P. debilis be studied with special focus on the role of polyphenolics and its broad utilization. The roots of P. debilis have more flavonoid contents than the aerial parts (Perera et al., 2016).

# iv) Organ protective effects

Among many other ailments, renal disease is considered as the 9th major cause of mortality across the world, and is considered the sole clinical sign of this disease. The disease is characterized by reduced excretory function by the kidney, and hence reduced glomerular filtration rate (GFR), with abnormal homeostasis in blood chemistry. Renal failure disease is mainly caused by environmental pollution, particularly by heavy metals which pose a threat to the biosphere and and particular to people exposed to industrial wastes or effluents, or farm workers, primarily attacking the kidneys. For instance, cadmium (Cd) enters the kidneys where it deposits in the proximal tubules (Olubunmi et al., 2017). Cadmium causes a decline in kidney activity by elevating blood urea and creatinine levels, which leads to further kidney damage by exposure to leaf extracts of P. amarus. It was suggested by Olubunmi et al. (2017) that P. amarus extracts have no prophylactic or ameliorative effects on Cdmediated kidney damage, and rather that continuous exposure to these extracts are deleterious to the kidney, but the Ayurvedic system of medication reports that P. emblica fruits are potent herbs, and hydroalcoholic extracts provide protection against hepatic disorders. Ethanolic extracts of fruits of P. emblica provide hepatoprotection mainly because of the high ROS scavenging and antioxidant activity of their main constituents like gallic acid and ellagic acid (Abbas et al., 2017; Chaphalkar et al., 2017). In different human diseases like diabetes, obesity and cardiovascular issues, insulin resistance is increased which is primarily due to higher intake of sugars. The ethanolic extracts of P. amarus are quiet beneficial and ameliorate insulin resistance by reducing metabolic syndrome and hepatic ROS levels in rats (Putakala et al., 2017). Ethanolic extracts of P. emblica L. dried fruits, have been reported to show positive gastroprotective effects in patients by reducing pain, vomiting, insomnia and sleeping disturbances or associated problems (Iqbal et al., 2017). Water extract of Р. emblica potentially reduced lipid peroxidation, mRNA expressions of CYP2E1, TNF and IL-1, while increasing antioxidant activity and reducing non-alcoholic steatohepatitis in vivo (Tung et al., 2018). However, although P. niruri extracts are enriched with anti-oxidant and antiinflammatory activities, no significant clinical benefits were observed in non-alcoholic steatohepatitis treatment (Hassan et al., 2017). The fruit extracts of P. emblica have ellagic acid as the major component, which helps reduce the ROS generation and fat accumulation. while modulating the expression of lipogenesis- linked genes and up-regulating AMPK signaling in HepG<sub>2</sub> cells. Extracts showed an inhibition in cellular steatosis and liver fibrosis in vitro (Lu et al., 2016).

v) Analgesic and anti-inflammatory activity

Inflammations are the immune responses to any disturbance in cells or tissues under stress. Prolonged exposure to these various stresses such as pathogens, chemicals, UVR, and pollutants. etc. may lead to serious inflammation that leads to chronic diseases. Inflammation and pain are quiet frequent causes of medical discussion which generally occurs when a tissue is injured (Wagle et al., 2016). Recently much focus has been devoted to screening and isolating novel drugs with analgesic potential from various plant sources in order to minimize pain or inflammation with less side effects compared toWestern medicines (Wagle et al., 2016). The members of the genus Phyllanthus are well studied for their analgesic and anti-inflammatory effects. In a recent study where Swiss albino mice were evaluated for the anti-inflammatory and analgesic properties of leaf ethanolic extracts of *P. acidus*, and it was observed that these extracts displayed remarkable action against inflammation and pains when applied at a final concentration of about 200 mg/kg of total body weight (Hossain et al., 2016). A comparative study to find anti-inflammatory. anti-arthritic and analgesic activity of various herbal extracts of Bacopa monnieriis, Cassia fistula and P. polyphyllus was conducted by Yoon and Lee (2017), which claimed that the extracts in various combination (w/w/w) = 1/2/1) had significant anti-inflammatory and analgesic actions. The aqueous extracts of the aerial parts of *P. muellerianus* along with its prominent secondary metabolites, e.g., geraniin, were found to have potential both peripheral and central anti-nociceptive effects in murine models of chemical nociception with the anti-nociceptive action of geraniin involving possibly the opioidergic pathways (Boakye-Gyasi et al., 2016). The green synthesis of silver nanoparticles (AgNPs) using aqueous fruit extracts of P. acidus represents an environmentally friendly and cheaper source of materials with potential therapeutic roles in cytoprotection and antiinflammation through scavenging nitric oxide and superoxide anions (Manikandan *et al.*, 2017). Furthermore, short-term exposure to *P. acidus*-mediated green-synthesized AgNPs did not affect the viability of peritoneal macrophages, and could be a potential therapeutic to treat inflammatory diseases by reducing the expression of IL-1 (Manikandan *et al.*, 2017).

The central nervous system (CNS) depressant measured along activity was with antidiarrheal and antipyretic activities of ethanolic leaf extract of P. acidus L., which reduced CNS depression significantly in animal models in a dose dependent way (Hossain et al., 2016). Moreover, P. acidus significant showed antidiarrheal and antipyretic actions, and therefore it could be an excellent source for natural CNS depressant, antidiarrhoeal and antipyretic agents for medical applications (Hossain et al., 2016). The comparative therapeutic efficacy of P. emblica fruits extract and procaine penicillin in the treatment of subclinical mastitis was studied in 30 subclinical mastitis positive buffaloes. It is concluded that P. emblica fruit extract is an inexpensive source to treat subclinical mastitis in dairy buffaloes and can be used as an alternative to antibiotic therapy like procaine penicillin (Khan et al., 2018). In inflammatory ulceration P. niruri is commonly applied in traditional medication systems to treat ulcers and inflammation. The methanolic extract of P. niruri leaves was used for its anti-inflammatiory and anti-ulcer activities, which suggests that leaf extracts are strong enough to reduce inflammation and provide protection against ulceration, as ascertained by regeneration of mucosal layer and substantially prevented hemorrhage and edema (Mostofa et al., 2017).

Another study recently evaluated the antinociceptive and anti-inflammatory potential of phytosterols isolated from the chloroform extracts of *P. maderaspatensis* (CEPM) through the carrageenan-induced hind paw oedema and hot plate method in male Wistar rats. CEPM extract and pentazocin had significant effects on the increase of the basal reaction time compared to control. This demonstrated the potential anti-inflammatory and analgesic effect of the CEPM which supports the claims by traditional medicine practitioners (Wagle *et al.*, 2016). Significant hypoglycemic, anti-diarrheal, analgesic, and anesthetic activities were shown by *P. acidus* pulp extracts (Afrin *et al.*, 2016).

## vi) Immunomodulatory effects

The human immune system is an organized system comprised of many different immune cells such as macrophages, neutrophils, Tlymphocytes, natural killer cells and various other specialized immune molecules that have resistance evolved to mediate against infections. Wide ranging ethnomedicinal or traditional uses of Phyllanthus plants are mainly linked to its active ingredients and pharmacological broad actions, e.g., immunomodulation, anti-viral and antibacterial, diuretic, anti-hyperglycemia and hepatoprotector properties (Tjandrawinata et al., 2017). Phytochemicals used as immunestimulants, are superior to conventional chemotherapeutics and antibiotics. Tuberculosis (TB) is a severe disease, and its progression mainly depends on the strength of the host immune system. P. niruri has been reported to boost the immune system in traditional medicine, and it is said that it has a great potential to induce immune cell activity in TB patients in vitro, The P. niruri extracts mainly help the release of nitric oxide and hence elevate the phagocytic activity of macrophages in a dose dependent fashion, ultimately modulating the immune responses (Putri et al., 2018). Moreover, P. niruri leaf extracts have immunostimulating effects on neutrophil activation and the antibody response of Oreochromis mossambicus. It is further demonstrated that P. niruri plant

extracts and their components can be used either as a routine feed supplement to activate the immune system of farmed fishes or as an adjuvant to enhance the efficacy of vaccines (Muthulakshmi et al., 2016). P. amarus strongly inhibits the phagocytic activity of human neutrophils and reduces cellular immune responses in rats. Moreover, the plant extracts have strong inhibitory effects on cellular and humoral immune responses (ceruloplasmin and lysozyme) suggesting the potential of the plant to be developed as an effective immunosuppressive agent which mainly acts through the inhibition of myeloperoxidase activity and nitric oxide release leading to the release of serum level immunoglobulins (Ilangkovan et al., 2016a). Phyllanthin, one of the main and active ingredient in many Phyllanthus species (P. amarus), has recently been reported to have hepatoprotective and immunosuppressive effects on various in Balb/C mice in a dosedependent way through the inhibition of CD11b/CD18 adhesion, nitric oxide and myeloperoxidase activity release (Ilangkovan et al., 2016a). Based on the level of doses of phyllanthin, a significant inhibition in the proliferation of B and T lymphocytes and down-regulation of Th1 (IL-2 and IFN-) and Th2 (IL-4) cytokines, CD4+ and CD8+ was noted (Ilangkovan et al., 2016a).

## vii) Antibacterial activities

The ability of a diverse class of microbes to resist known anti-microbial drugs is always been a universal problem leading to exploration of novel anti-microbial drugs from diverse natural sources such as medicinal plants, especially the medicinal flora which has not been examined for their pharmacological potential as anti-bacterial potentials. *Phyllanthus niruri* L. is one of the plants that can be used as a preventive and alternative treatment as a substitute for antibiotics for the treatment of Chronic Respiratory Disease (CRD) in broiler chickens caused by Mycoplasma galisepticum. The chemicals contained in meniran (P. niruri L.) include antibacterial tannins, saponins, and alkaloids. A 30% plant extract caused up to 65% growth inhibition in Mvcoplasma galisepticum (Sabdoningrum et al., 2017). In continuation of a similar study, Ramandeep and colleagues found that P. niruri extracts could potentially inhibit the growth of Escherichia coli, Lactobacillus acidophilus, Pseudomonas aeruginosa and Staphylococcus aureus. Moreover, P. niruri is rich in phytochemicals that have antioxidant and antimicrobial activity, suggesting the need for further in-depth investigations for its systematic use in traditional medicine (Lee et al., 2016).

The extracts from P. amarus leaves were examined for complete chemical analyses along with their anti-microbial potential. Various stable extracts, i.e., hexane (59%), acetone (57%), and water extracts (48%) were prepared, which contain many alkaloids, saponins, anthraquinones, tannins and phenolics in acidic medium. In general, combining leaf extracts with either bacitracin or erythromycin alone has synergistic effects, thus depicting the crucial advantage of such combinations of these universal antibiotics with extracts of *P. amarus* to treat various infections (Senjobi et al., 2017). Various other studies examined the anti-microbial activities for aqueous extracts of above ground parts of P. muellerianus, and it was stated that geraniin is the major component in these extracts. In certain an agar-well diffusion assay, micro-dilution method, and time-kill kinetic studies were applied to find the anti-microbial potential of areal plant parts geraniin have where strong extracts. inhibitory effects on E. coli, P. aeruginosa, S. aureus, B. subtilis, and two clinical isolates viz. S. pyogenes and C. albicans, and significantly inhibited the zones of growth. Moreover. the minimal inhibitory

concentrations of these aerial plant parts extracts and geraniin were in a variable ranges i.e., 0.31 to 5 and 0.08 to 1.25 mg/mL, respectively. In another experiment where minimum cidal concentration were recorded for the plant extracts (5.0 to 50.0) and geraniin (2.5 to 10 mg/mL), respectively, thus proven that both the extracts and geraniin have strong anti-microbial potential. All these initial studies regarding in-depth exploration of various chemical constituents in P. muellerianus extracts further confirmed the occurrence of various terpenes, flavones, saponins, tannins, alkaloids and glycosides. It is further said that, these extracts have geraniin as primer constituent which pose strong anti-microbial potential against the above mentioned bacterial strains (Boakye et al., 2016).

In continuation of similar finding, a study examined the antimicrobial potential of volatile oils extracted from leaves of P. where they found good muellerianus, inhibitory effects partly on the isolated pure extracellular protease of pathogenic Klebsiella granulomatis. These volatile oils showed strong antimicrobial potential as depicted by their inhibitory capability against these proteases. Further the authors proposed that in-depth studies and clinical trials using these oils could define their effectiveness to combat various skin conditions such as third degree burns, cuts, injuries and post-operative wounds infections caused by K. granulomatis as well as many related strains (Adesegun et al., 2016). Pathmavathi and Thamizhiniyan (2016) prepared various leaf extracts (methanolic, hexane, chloroform, and ethyl acetate) of P. amarus and Plectranthus ambionicus to study their antifungal and antimicrobial potential. Four Gram positive and three Gram negative bacterial strains along with three species of fungi were tested to screen the best isolate among the various organic plant extracts. The anti-microbial

activity of various extracts of both these plant species showed mixed levels of anti-microbial potential towards the selected fungal and bacterial strains, but the ethyl acetate extracts of Plectranthus ambionicus and P. amarus showed superior antimicrobial potential compared to other extracts. Recently zinc sulphide nanoparticles were synthesized from P. emblica leaves and fruit extract to analyze the activity of these significant phytochemicals. These zinc sulphide nanoparticles possess potential antimicrobial activity against many pathogenic organisms (Sathishkumar et al., 2017). The anti-Helicobacter pylori and urease inhibitory activity of hydroalcoholic extracts of P. niruri L. was studied, and it was found that quercetin is one of the major constituents which is thought to be the cause of noncompetitive urease inhibition (Kaur et al., 2016). The leaf extracts of P. amarus showed inhibitory effects on Streptococcus pyrogenes, Streptococcus pneumonae, S. aureus, P. aeruginosa and Candida albicans in a study by Oluboyo et al. (2016). Many of the phytochemicals (flavones, terpenoides, various alkaloids. benzenoids, steroids. saponins and complex lipid molecules) were compared with ampicillin, gentamicin and observed pefloxacin, and to possess significant inhibition activity against the growth of selected studied microorganisms in a dose dependent fashion. Moreover, well antibiotics known had no significant inhibitory effect on the tested microorganisms compared to the leaf extracts of *P. amarus* (Oluboyo et al., 2016). Gunawan et al. (2016) have reported that *P. niruri* extracts mainly contain alkaloids, triterpenoids and flavones, which are highly efficcient to treat dysentery, rheumatism, inflammation and gut worms in children in traditional medication systems. Furthermore, active triterpenoids in the chloroform extracts from the bark of *P. niruri* possess strong antibacterial activity and control the proliferation of S. aureus at an

optimum concentration (1000 ppm) evidenced by a great 12 mm inhibition zone (Gunawan *et al.*, 2016). In another study, remarkable anti-bacterial potential against various fungal and bacterial strains was seen by using silver nanoparticles (AgNPs) obtained from a supercritical CO2 extract of *P. niruri* (Haris *et al.*, 2017). The phytochemical analyses of *P. amarus* leaf extracts were studied and presence of alkaloids, saponins, tannins, flavonoids, cyanogenic glycosides and steroids for antimicrobial activity.

The phytochemical examination of the leaf extract of *P. amarus* shows the presence of a variety of bioactive components: alkaloids, saponins, tannins flavonoids, cyanogenic glycosides and steroids in ethanol and ethyl acetate. The extracts of both these solvents were applied to find their potential as antimicrobial agents against Staphilococcus aureus, Escherichia coli and Clostridium albicans, and were found pretty suitable for the production of advanced anti-microbial drugs (Uzor et al., 2016). The leaf methanolic extracts of P. niruri showed a well defined antimicrobial activity against Coney lunata and Salmonella typhi, and it was suggested by the authors that this plant is of great clinical importance and may potentially be used in the pharmaceutical industry (Ahamath et al., 2017). Chewing gums were prepared having constituents of fruit extract of P. emblica and studied in detail for their potential in alleviating and soothing oral health problems (Gao et al., 2018). Fruit extracts of P. emblica have great pharmacological potential against bacterial growth, inflammation, cellular oxidative stress induced by ROS, and various types of cancers as well as different oral diseases. The focus of the study was changes in the oral microflora in a gum-basecontrolled crossover manner caused by the effect of *P. emblica* leaf extracts. Moreover, chewing gums prepared with the extracts of Phyllanthus fruits help to stimulate saliva production, producing a significant reduction in the recurrence of clinical oral microflora for a short time period, and are thus proven as a safer way of improving oral health (Gao *et al.*, 2018).

## Conclusion

The medicinal potentials of these herbs have been documented over some decades. There is therefore, the need to explore its integration in standardized form into alternative medicine and pharmaceutical industries to cure an array of diseases in man and animals. It could also be harnessed as a replacement for the hitherto antibiotic growth promoters in production of animals since the conventional synthetic antibiotics have been banned in most countries of the world due to antibiotic resistance they confer on recipient animals and the end users; humans. Therefore, this compilation will aid further research into the veracity of claims that these plants contain phytochemicals that aid cure/prevention of some diseases.

## References

Abbas N, Naz M, Alyousef L, Ahmed ES, Begum A (2017). Comparative study of hepatoprotective effect produced by Cuminum cyminum, fruits of *Phyllanthus emblicus* and silymarin against cisplatininduced hepatotoxicity. International Journal of Pharmaceutical Sciences Research 8, 2026–2032.

Adedapo AA, Adegbayibi AY, Emikpe BO (2005). Some clinico-pathological changes associated with the aqueous extract of the leaves of *Phyllanthus amarus* in rats. Phytotherapy Research 19, 971–976.

Adeneye AA, Senebo AB (2008). Protective effect of aqueous leaf and seed extract of *Phyllanthus amarus* on gentamicin and acetaminophen induced nephrotoxic rats. Journal of Ethnopharmacology 111, 318-323.

Adeolu AA, Sunday OO (2013). Antiinflammatory and analgesics activities of soft drink extract of *Phyllanthus amarus* in some laboratory animals. British Biotechnology Journal 3, 191-204.

Adesegun A, Samuel F, Adesina O (2016). Antibacterial activity of the volatile oil of *Phyllanthus muellerianus* and its inhibition against the extracellular protease of *Klebsiella granulomatis*. European journal of medicinal plants 14 (2), 1-10.

Afrin F, Banik S, Hossain MS (2016). Pharmacological activities of methanol extract of *Phyllanthus acidus* pulp. Journal of Medicinal Plants Research 10, 790–795.

Ahamath JM, Vahith RA, Manivel V (2017). Elamparithi, R. Phytochemical Screening and Antimicrobial Activity of *Phyllanthus niruri*. Journal of Advances in Applied Science Research 1, 7-11.

Akporowhe S, Onyesom I (2016). Phyllanthus amarus augments the serum antioxidant capacity and invigorates the blood in experimental mice. Bioscience Biotechnolology Research. Communications 9, 15–18.

Andrianto D, Widianti W, Bintang M (2017). Antioxidant and Cytotoxic Activity of *Phyllanthus acidus* Fruit Extracts. IOP Conference Series: Earth and Environment Science 58, 012-022.

Angamuthu J, Ganapathy M, Evanjelene VK (2016). Evaluation of antioxidant activity of *Phyllanthus acidus*. World Journal Pharmacy and Pharmaceutical Sciences 5, 1011–1016.

Bagalkotkar G, Sagineedu SR, Saad MS, Stanslas J (2006). Phytochemicals from *Phyllanthus niruri* Linn. and their pharmacological properties: a review. Journal of Pharmacy and Pharmacology 58, 1559– 1570. doi: 10.1211/jpp.58.12.0001

Bahar L, Sarker SD, Delazar A (2011). "Phytochemistry of the genus *Phyllanthus*," in *Phyllanthus* species Scientific Evaluation and Medicinal Applications, R. Kuttan and K.B. Harikumar, Eds., pp. 119-138.

Balakrishnan NP, Chakrabarty T (2007). The family Euphorbiaceae in India; a synopsis of its profile, taxonomy and bibliography (Euphorb India). Dehra Dun, India: Bishen Singh Mahendra Pal Singh.

Bekoe EO, Kitcher C, Debrah P, Amoateng P, Donkor PO, Martinson. A (2020). Study on *Phyllanthus amarus*; Pharmacognostic, Mycobactericidal and Mutagenic Properties. Pharmacognosy Journal 12(6)Suppl, 1732-9.

Boakye YD, Agyare C, Dapaah SO (2016). In vitro and in vivo antioxidant properties of *Phyllanthus muellerianus* and its major constituent, geraniin. Oxidants and Antioxidants in Medical Science 5, 70–78.

Boakye-Gyasi E, Kasange EA, Biney RP, Boadu-Mensah K, Agyare C, Woode E (2016). Anti-nociceptive effects of geraniin and an aqueous extract of the aerial parts of *Phyllanthus muellerianus* (Kuntze) Exell. in murine models of chemical nociception. Iranian Journal Pharmaceutical Sciences 12, 17–30.

Borges LDC, Negrão-Neto R, Pamplona S, Fernandes L, Barros M,; Fontes-Júnior E, Maia C, Silva CYY, Silva MND (2018). Anti-Inflammatory and Antinociceptive Studies of Hydroalcoholic Extract from the Leaves of *Phyllanthus brasiliensis* (Aubl.) Poir. and Isolation of 5-O-\_-D-Glucopyranosyljusticidin B and Six Other Lignans. Molecules 23, 941-946.

JCBR Vol 3 Is 4 July-Aug 2023

Burkill H (1966). A dictionary of the economic products of the Malay Peninsula. 2nd Ed. Vol. 2. Kuala Lumpur, Malaysia: Ministry of Agriculture & Co-operatives, p. 2444.

Calixto JB, Santos ARS, Filho VC, Yunes RA (1998). A review of the plants of the genus *Phyllanthus*: Their chemistry, pharmacology, and therapeutic potential. Medicinal Research Reviews 18, 225–258. doi: 10.1002/(SICI)1098-1128(199807)18:4<225::AID-MED2>3.0.CO;2-X

Chansriniyom C, Bunwatcharaphansakun P, Eaknai W, Nalinratana N, Ratanawong A, Khongkow M, Luechapudiporn R (2018). A synergistic combination of *Phyllanthus emblica* and *Alpinia galanga* against H2O2induced oxidative stress and lipid peroxidation in human ECV304 cells. Journal of Functional Foods 43, 44–54.

Chaphalkar R, Apte KG, Talekar Y, Ojha SK, Nandave M (2017). Antioxidants of *Phyllanthus emblica* L. bark extract provide hepatoprotection against ethanol-induced hepatic damage: A comparison with silymarin. Oxidative Medicine and Cellular Longevity 2017, 3876040.

Chung CY, Li CH, Burnouf T, Wang GH, Chang SP, Jassey A, Tai CJ, Tai CJ, Huang CJ, Richardson CD (2016). Activity-based and fraction-guided analysis of *Phyllanthus urinaria* identifies loliolide as a potent inhibitor of hepatitis C. virus entry. Antiviral Research 130, 58–68.

Dada EO, Ekundayo FO, Makanjuola OO. (2014). Antibacteria activities of *Jatropha curcas* Linn on coliforms isolated from surface waters in Akure, Nigeria. International Journal of Biomedical Science 10(1), 25-30.

Dai M, Wahyuni A, Dk I, Azizah T, Suhendi A, Saifudin A (2016). Antioxidant activity of *Phyllanthus niruri* L. herbs: In vitro and in vivo models and isolation of active compound. National Journal of Physiology, Pharmacy and Pharmacology 6, 32–37.

De Oliveira CNF, Frezza TF, Garcia VL, Figueira GM, Mendes TMF, Allegretti SM (2017). Schistosoma mansoni: In vivo evaluation of *Phyllanthus amarus* hexanic and ethanolic extracts. Experimental Parasitology 183, 56–63.

Devi S, Rashid R, Kumar M (2017). Phytochemistry and pharmacological properties of *Phyllanthus amarus* schum: a review. The Pharma Innovation Journal 6 (12), 169-172.

Devi S, Kumar D, Kumar M (2016). In-Vitro antioxidant activities of methanolic extract of whole plant of *Phyllanthus amarus* (Euphorbiaceae). International Journal of Botany Studies 1, 30–32.

Etta H (2008). Effects of *Phyllanthus amarus* on litter traits in albino rats. Scientific Research and Essay, 3(8), 370-372.

Evi PL, Degbeku K (2011). Antidiabetic activity of *Phyllanthus amarus* schum and thonn and alloxan induced diabetes in male Wistar rats. Journal of Applied Sciences 11(6), 2968-2973.

Fan YY, Gan LS, Liu HC, Li H, Xu CH, Zuo JP, Ding J, Yue JM (2017). Phainanolide, A. Highly modified and oxygenated triterpenoid from *Phyllanthus hainanensis*. Organic Letters 19, 4580–4583.

Foo LY (1995). Amarinic acid and related ellagitannins from *Phyllanthus amarus*. Phytochemistry 39, 217–224. doi: 10.1016/0031-9422(94)00836-I

Foo LY, Wong H (1992). Phyllanththusiin D, an unusual hydrolyzable tannin from *Phyllanthus amarus*. Phytochemistry 31, 711– 713. doi: 10.1016/0031-9422(92)90071-W

Foo LY (1993). Amariin a di-dehydro hexahydroxy diphenoyl hydrolysable tannin from *Phyllanthus amarus*. Phytochemistry 33, 487–491.

Gao Q, Li X, Huang H, Guan Y, Mi Q, Yao J (2018). The Efficacy of a Chewing Gum Containing *Phyllanthus emblica* Fruit Extract in Improving Oral Health. Current Microbiology 75, 604–610.

Gunawan I, Bawa I, Putra AB (2016). Isolation, characterization and antibacterial activity of triterpenoid compounds fraction chloroform bark *Phyllanthus niruri* L. World Journal of Pharmacy and Pharmaceutical Science 5, 357–364.

Haris M, Kumar A, Ahmad A, Abuzinadah MF, Basheikh M, Khan SA, Mujeeb M (2017). Microwave-assisted green synthesis and antimicrobial activity of silver nanoparticles derived from a supercritical carbon dioxide extract of the fresh aerial parts of *Phyllanthus niruri* L. Tropical Journal of Pharmaceutical Research 16, 2967–2976.

Hassan MRA, Mustapha NRN, Jaya F, Arjunan S, Ooi ET, Said RM, Menon J, Tee HP, Omar H, Aiman S (2017). Efficacy and Safety of *Phyllanthus niruri* in Non-alcoholic Steatohepatitis Treatment: Pilot Study from Malaysia. Journal of Pharmacy Practice and Community Medicine, 3, 131–137. Hidanah S, Sabdoningrum EK, Rachmawati K, Soeharsono S, Trika GGA, Widiati TP (2022). The activity of Meniran (*Phyllanthus niruri* Linn.) extract on Salmonella pullorum infected broilers. Veterinary World, 15(5), 1373-1382. doi: 10.14202/vetworld. 2022.1373-1382

Hidanah S, Sabdoningrum EK, Wahjuni RS, Chusniati S (2018). Effects of meniran (*Phyllanthus niruri L.*) administration on leukocyte profile of broiler chickens infected with *Mycoplasma gallisepticum*. Veterinary World, 11(6), 834-839. doi: 10.14202/vetworld.2018.834-839

Hoffmann P, Kathriarachchi H, Wurdack KJ (2006). A phylogenetic classification of Phyllanthaceae (Malpighiales; Euphorbiaceae sensu lato). Kew Bulletin 61 (1), 37–53.

Hossain M, Akter S, Das A, Sarwar M (2016). CNS Depressant, Antidiarrheal and Antipyretic Activities of Ethanolic Leaf Extract of *Phyllanthus acidus* L. on Swiss Albino Mice. British Journal of Pharmaceutical Research 10, 1–9.

Houghton PJ, Woldemariama TZ, Siobhan OS, Thyagarajan SP (1996). Two securinega type alkaloids from *Phyllanthus amarus*. Phytochemistry 43, 715–717.

Huang ST, Huang CC, Sheen JM, Lin TK, Liao PL, Huang WL, Wang PW, Liou CW, Chuang JH (2016). *Phyllanthus urinaria*'s Inhibition of Human Osteosarcoma Xenografts Growth in Mice is Associated with Modulation of Mitochondrial Fission/Fusion Machinery. The American Journal of Chinese Medicine 44, 1507–1523.

Ifeoma O, Samuel O, Itohan AM, Adeola SO (2013). Isolation, fractionation and evaluation of the antiplasmodial properties of *Phyllanthus niruri* resident in its

JCBR Vol 3 Is 4 July-Aug 2023

chloroform fraction. Asian Pacific Journal of Tropical Medicine 6, 169–175. doi: 10.1016/S1995-7645(13)60018-8

Iizuka T, Nagai M, Taniguchi A, Moriyama H, Hoshi K (2007). Inhibitory Effects of Methyl Brevifolincarboxylate Isolated from *Phyllanthus niruri L*. on Platelet Aggregation. Biological and Pharmaceutical Bulletin 30(2), 382-384.

Ilangkovan M, Jantan I, Bukhari SNA (2016). Phyllanthin from *Phyllanthus amarus* inhibits cellular and humoral immune responses in Balb/C mice. Phytomedicine, 23: 1441–1450. doi: 10.1016/j.phymed.2016.08.002

Iqbal Z, Asif M, Aslam N, Akhtar N, Asmawi MZ, Fei YM, Jabeen Q (2017). Clinical investigations on gastroprotective effects of ethanolic extract of *Phyllanthus emblica* L. fruits. Journal of Herbal Medicine 7, 11–17.

Ishimaru K, Yoshimatsu K, Yamakawa T, Kamada H, Shimomura K (1992). Phenolic constituents in Tissue Cultures of *Phyllanthus niruri*. Phytochemistry, 31(6): 2015-2018.

Jain N, Kumar AS, Shefali S, Preet SSK, Kumar S (2008). SCAR Markers for Correct Identification of *Phyllanthus amarus*, *P. fraternus*, *P. debilis* and *P. urinaria* used in Scientific Investigations and Dry Leaf Bulk Herb Trade. Planta Medica 74, 296-301.

Jantan I, Ilangkovan M, Mohamad HF (2014). Correlation between the major components of Phyllanthus amarus and Phyllanthus urinaria and their inhibitory effects on of phagocytic activity human neutrophils. BMC Complementary Alternative Medicine 14. 429. doi: 10.1186/1472-6882-14-429

Kalpana S, Ramakrushna B, Anitha S (2016). Evaluation of in vitro antioxidant and amylase inhibitory activity of *Phyllanthus indofischeri* Bennet. International Journal of Pharmacy and Pharmaceutical Sciences 8, 131–136.

Kassuya CA, Silvestre A, Menezes-de-Lima O, Marotta DM, Rehder VLG, Calixto JB (2006). Antiinflammatory and antiallodynic actions of the lignan niranthin isolated from *Phyllanthus amarus*: evidence for interaction with platelet activating factor European receptor. Journal of Pharmacology 546, 182–188. doi: 10.1016/j.ejphar.2006.07.025

Kaur B, Kaur N, Gautam V (2016). Evaluation of anti-helicobacter pylori (DSMZ 10242) activity and qualitative analysis of quercetin by HPLC in Phyllanthus niruri linn. World Journal of Pharmacy and Pharmaceutical Sciences 5, 1691–1706.

Kawakita A, Kato M (2009). Repeated independent evolution of obligate pollination mutualism in the Phyllantheae-Epicephala association. Proceedings of the Royal Society B 276, 417–426. doi: 10.1098/rspb.2008.1226

Khan A, Ahmed T, Rizwan M, Khan N (2018). Comparative therapeutic efficacy of *Phyllanthus emblica* (Amla) fruit extract and procaine penicillin in the treatment of subclinical mastitis in dairy buffaloes. Microbial Pathogenesis 115, 8–11.

Komlaga G, Genta-Jouve G, Cojean S, Dickson RA, Mensah ML, Loiseau PM, Champy P, Beniddir MA (2017). Antiplasmodial Securinega alkaloids from *Phyllanthus fraternus*: Discovery of natural (+)-allonorsecurinine. Tetrahedron Letters 58, 3754–3756.

Kuttan R, Harikumar KB (2011). "Phyllanthus Species," in Scientific Evaluation and Medicinal Applications, Edition 1st (Boca Raton: CRC Press), 388. Edition 2011 eBook Published 29 August 2011. doi: 10.1201/b11380

Lee NY, Khoo WK, Adnan MA, Mahalingam TP, Fernandez AR. Jeevaratnam K (2016). The pharmacological potential of *Phyllanthus niruri*. Journal of Pharmacy and Pharmacology 68, 953–969.

Lee SK, Li PT, Lau DT, Yung PP, Kong RY, Fong WF (2006). Phylogeny of medicinal *Phyllanthus* species in China based on nuclear ITS and chloroplast atpB-rbcL sequences and multiplex PCR detection assay analysis. Planta Medica 72, 721–726. doi: 10.1055/s-2006-931580

Leite DF, Kassuya CA, Mazzuco TL, Silvestre A, De-Melo LV, Rehder VL (2006). The cytotoxic effect and the multidrug resistance reversing action of lignans from *Phyllanthus amarus*. Planta Medica 72, 1353–1358.

Lestariningsih SO, Sudjarwo E (2015a). Pengaruh tepung tanaman meniran terhadap aktivitas antimikroba bakteri asam laktat dan *Escherichia coli*. Jurnal Ilmu-Ilmu Peternakan 25, 55-60.

Lestariningsih SO, Sudjarwo E (2015b). Pengaruh tepung tanaman meniran (*Phyllanthus niruri* Linn) sebagai pakan tambahan terhadap mikroflora usus halus ayam pedaging. Jurnal Agripet 15, 85-91.

Lim Y, Murtijaya J (2007). Antioxidant properties of *Phyllanthus amarus* extract as affected by different drying methods. Food Science and Technology 40(9), 1664-1669.

Londhe JS, Devasagayam TP, Foo LY, Ghaskadbi SS (2009). Radioprotective properties of polyphenols from *Phyllanthus amarus* Linn. Journal of Radiation Research, 50, 303–309.

Lu CC, Yang SH, Hsia SM, Wu CH, Yen GC (2016). Inhibitory effects of *Phyllanthus emblica* L. on hepatic steatosis and liver fibrosis in vitro. Journal of Functional Foods 20, 20–30.

Maciel MAM, Cunha A, Dantas FTNC, Kaiser CR (2007). NMR characterization of bioactive lignans from *Phyllanthus amarus* Schum & Thonn. Journal of Magnetic Resonance Imaging 6, 76–82.

Manikandan R, Beulaja M, Thiagarajan R, Palanisamy S, Goutham G, Koodalingam A, Prabhu NM, Kannapiran E, Basu MJ, Arulvasu C (2017). Biosynthesis of silver nanoparticles using aqueous extract of *Phyllanthus acidus* L. fruits and characterization of its anti-inflammatory effect against  $H_2O_2$  exposed rat peritoneal macrophages. Process Biochemistry 55, 172– 181.

Manikkoth S, Dcepa B, Joy AE, Rao S. (2011). Anti-convulsant activity of *Phyllanthus amarus* in experimental animal models 4, 144-149.

Mao X, Wu LF, Guo HL, Chen WJ, Cui YP., Qi, Q., Li, S., Liang, W.Y., Yang, G.H., Shao, Y.Y., Zhu, D. and She, G.M. (2016) The genus *Phyllanthus* : an ethnopharmacological, phytochemical, and pharmacological review. Evid Based Complement Alternat Med., volume 2016, Article ID 7584952, 36 pages, https://doi.org/10.1155/2016/7584952

Maria AM, Gineide CA, Sthephanye MR, Deborah DP, Heryka MMMR (2019). Botanic, Phytochemistry and Pharmacological Aspects of *Phyllanthus amarus* Schum. & Thonn. as Powerful Tools to Improve its Biotechnological Studies. Ann. Chem. Sci. Res., 1(2): 1-9. DOI: 10.31031/ACSR.2019.01.000510

JCBR Vol 3 Is 4 July-Aug 2023

Mediani A, Abas F, Maulidiani M, Khatib A, Tan CP, Ismail IS, Shaari K, Ismail A (2017). Characterization of metabolite profile in *Phyllanthus niruri* and correlation with bioactivity elucidated by nuclear magnetic resonance based metabolomics. Molecules. 22, 1-14. doi:10.3390/molecules22060902

Meena J, Sharma RA, Rolania R (2018) A review on phytochemical and pharmacological properties of *Phyllanthus amarus* schum and thonn. International Journal of Pharmaceutical Sciences and Research 4(9): 1377-1386.

Menéndez-Perdomo IM, Wong-Guerra M, Fuentes-León F, Carrazana E, Casadelvalle I, Vidal A, Sánchez-Lamar Á (2017). Antioxidant, photoprotective and antimutagenic properties of *Phyllanthus* spp. From Cuban flora. Journal of Pharmacy and Pharmacognosy Research 5, 251–261.

Mitra RI, Jain SK (1985). Concept of *Phyllanthus niruri* (Euphorbiaceae) in Indian Floras. Bulletin of the Botanical Survey of India, 27, 161–176.

Moronkola DO, Ogunwande IA, Oyewole IO, Baser KHC, Ozek T, Ozek G (2009). Studies on the volatile oils of *Momordica charantia* L (Cucurbitaceae) *Phyllanthus amarus* Sch and Thonn (Euphorbiaceae). Journal of Essential Oil Research 21, 393–399.

Mostofa R, Ahmed S, Begum MM, Sohanur Rahman M, Begum T, Ahmed SU, Tuhin RH, Das M, Hossain A, Sharma M (2017). Evaluation of anti-inflammatory and gastric anti-ulcer activity of *Phyllanthus niruri* L. (Euphorbiaceae) leaves in experimental rats. BMC Complementary and Alternative Medicine 17, 267-272. Mujeeb F, Bajpai P, and Pathak N (2014). Phytochemical evaluation, antimicrobial activity, and determination of bioactive components from leaves of *Aegle marmelos*. BioMedical Research International 2014, 1-14. doi.org/10.1155/2014/497606

Muthulakshmi M, Subramani P, Michael R (2016). Immunostimulatory effect of the aqueous leaf extract of *Phyllanthus niruri* on the specific and nonspecific immune responses of *Oreochromis mossambicus* Peters. Iranian Journal Veterinary Research 17, 200–202.

Muthusamy A, Prasad HNN, Sanjay ER, Rao MR, Satyamoorthy K (2016). Impact of precursors and plant growth regulators on in vitro growth, bioactive lignans, and antioxidant content of *Phyllanthus* species. In Vitro Cellular and Developmental Biology, 52, 598–607.

Nahar L, Sarker, SD, Delazar A (2011). "Phytochemistry of the genus *Phyllanthus*" in *Phyllanthus Species: Scientific Evaluation and Medicinal Applications*. Eds. Kuttan, R., Harikumar, K. B. (London, UK: Taylor and Francis Group, CRC Press), p. 119–138. doi: 10.1201/b11380-7

Nain P, Saini V, Sharma S, Nain J (2012). Antidiabetic and antioxidant potential of Emblica officinalis Gaertn. leaves extract in streptozotocin-induced type-2 diabetes mellitus (T2DM) rats. Journal of Ethnopharmacology 142, 65-71. doi: 10.1016/j.jep.2012.04.014

Nguyen GC, Smalley WE, Vege SS (2016). AGA Clinical Guidelines Committee. Gastroenterological Association American medical Institute guideline on the management microscopic colitis. of Gastroenterology 150, 242–246.

Nguyen VT, Sakoff JA, Scarlett CJ (2017). Physicochemical Properties, Antioxidant and Cytotoxic Activities of Crude Extracts and Fractions from *Phyllanthus amarus*. Medicines 4, 42-47.

Noorudheen N, Chandrasekharan DK (2016). Effect of ethanolic extract of *Phyllanthus emblica* on captan induced oxidative stress in vivo. South Indian Journal of Biological Sciences 2, 95–102.

Obianime AW, Uche FI (2009). The Phytochemical constituents and the effects of methanol extracts of *Phyllanthus amarus* leaves (kidney stone plant) on the hormonal parameters of Male guinea pigs. Journal of Applied Sciences and Environmental Management 13(1), 5-9.

Ojezele MO, Moke EG, Onyesom I (2017). Impact of generic antimalarial or *Phyllanthus amarus* and vitamin co-administration on antioxidant status of experimental mice infested with *Plasmodium berghei*. Beni-Suef University Journal of Basic Applied Sciences 6, 260–265.

Oluboyo BO, Oluboyo AO, Kalu SO (2016). Inhibitory effects of *Phyllanthus amarus* extracts on the growth of some pathogenic microorganisms. African Journal of Clinical and Experimental Microbiology 17, 166–172.

Olubunmi OP, Yinka OS, Oladele OJ, John OA, Boluwatife BD, Oluseyi FS (2017). Aberrations in Renal Function Parameters Following Oral Administration of *Phyllanthus amarus* in Cadmium-Induced Kidney Damage in Adult Wistar Rats. Journal of Diseases and Medicinal Plants 3, 60–67.

Omulokoli E, Khan B, Chhabra SC (1997). Antiplasmodial activity of four Kenyan medicinal plants. Journal of Ethnopharmacology 56, 133–137. doi: 10.1016/S0378-8741(97)01521-3

Patel JR, Tripathi P, Sharma V, Chauhan NS, Dixit VK (2011). *Phyllanthus amarus*: ethnomedicinal uses, phytochemistry and pharmacology: a review. Journal of Ethnopharmacology 138, 286–313. doi: 10.1016/j.jep.2011.09.040

Pathmavathi M, Thamizhiniyan P (2016). Antimicrobial activity of various extracts of *Plectranthus ambionicus* and *Phyllanthus amarus*. Journal of Applied and Advanced Research 1, 29–35.

Pereira RG, Garcia VL, Rodrigues MVN, Martínez J (2016). Extraction of lignans from *Phyllanthus amarus* Schum. & Thonn using pressurized liquids and low pressure methods. Separation and Purification Technology 158, 204–211.

Pereira RG, Nakamura RN, Rodrigues MVN, Osorio-Tobón JF, Garcia VL, Martinez J (2017). Supercritical fluid extraction of phyllanthin and niranthin from *Phyllanthus amarus* Schum. and Thonn. The Journal of Supercritical Fluids 127, 23–32.

Perera D, Soysa P, Wijeratne S (2016). Polyphenols contribute to the antioxidant and antiproliferative activity of *Phyllanthus debilis* plant in-vitro. BMC Complementary and Alternative Medicine 16, 339.

Peter S, Dey S, Veerakyathappa B, Kumar SR, Paulad C (2016). Therapeutic activity of partially purified fractions of *Emblica officinalis* (Syn. *Phyllanthus emblica*) dried fruits against *Trypanosoma evansi*. Journal of Pharmacy and Pharmacology 4: 546-558.

Prasad PD, Kavimani S, Suba V, Nudu T, Sanatorium T (2013). Antimicrobial activity

of the root extracts of *Phyllanthus amarus*. Applied Journal of Hygiene 4(1), 1039-1043.

Putakala M, Gujjala S, Nukala S, Bongu SBR, Chintakunta N, Desireddy S (2017). Cardioprotective effect of *Phyllanthus amarus* against high fructose diet induced myocardial and aortic stress in rat model. Biomedical Pharmacotherapy 95, 1359–1368.

Putri DU, Rintiswati N, Soesatyo MH, Haryana SM (2018). Immune modulation properties of herbal plant leaves: *Phyllanthus niruri* aqueous extract on immune cells of tuberculosis patient-*in vitro* study. Natural Product Research 32, 463–467. doi: 10.1080/14786419.2017.1311888

Rajeshkumar NV, Joy NV, Kuttan G, Nair MJ, Kuttan R (2002). Antitumor and anticarcinogenic activity of *Phyllanthus amarus* extract. Journal of Ethnopharmacology 81(1):17-22.

Ramandeep K, Nahid A, Neelabh C, Navneet K (2017). Phytochemical Screening of *Phyllanthus niruri* collected from Kerala Region and its Antioxidant and Antimicrobial Potentials. Journal of Pharmaceutical Sciences and Research 9, 1312–1316.

Rusmana D, Wahyudianingsih R, Elisabeth M, Balqis B, Maesaroh M, Widowati W (2017). Antioxidant Activity of *Phyllanthus niruri* Extract, Rutin and Quercetin. Indonesia Biomedical Journal 9, 84–90.

Sabdoningrum EK, Hidanah S, Wahjuni RS, Chusniati S, Arimbi A (2017). An in vitro antibacterial activity test of Meniran Herbs' (*Phyllanthus Niruri* L.) ethanol extract against *Mycoplasma gallisepticum* causes Chronic Respiratory Disease (CRD) in Broiler Chickens. KnE Life Sciences 3, 48–61. Sabir SM, Shah RH, Shah AH (2017). Total Phenolic and Ascorbic Acid Contents and Antioxidant Activities of Twelve Different Ecotypes of *Phyllanthus emblica* from Pakistan. Chiang Mai Journal of Science 44, 904–911.

Salazar JR, Martinez-Vazquez M, Cespedes CL, Ramirez-Apan T, Nieto-Camacho A, Flores Murrieta F (2011). Antiinflammatory and cytotoxic activities of Chichipegnin, peniocerol and macdougallin isolated from *Myrtillocactus geometrizans* Con. Zeitschrift fur Naturforschung C 66(1-2), 24-30.

Sangeeta D, Romana R, Muneesh K (2017). Phytochemistry anf pharmacological properties of *Phyllanthus amarus* Schum: *A review*. The Pharmaceutical Innovation Journal 6(12), 169-172.

Sarin B, Verma N, Martín JP, Mohanty A (2014). An overview of important ethnomedicinal herbs of *Phyllanthus* species: present status and future prospects. Scientific World Journal 2014, 12. doi: 10.1155/2014/839172

Sathishkumar M, Saroja M, Venkatachalam M, Rajamanickam A (2017). Biosynthesis of zinc sulphide nanoparticles using *Phyllanthus emblica* and their antimicrobial activities. Elixir Electrical Engineering 102, 44411–44415.

Senjobi CT, Ettu AO, Otujo CO (2017). Antibacterial and antifungal activities of leaf extracts of *Phyllanthus amarus* Schum and Thonn. Journal of Pharmacogosy and. Phytotherapy 9, 6–10.

Singh RP, Pal A, Pal K (2016). Antioxidant activity of ethanolic and aqueous extract of *Phyllanthus niruri*—In vitro. World Journal of Pharmacy and Pharmaceutical Science 5, 1994–2000.

Srivastava V, Singh M, Malasoni R, Shanker K, Verma RK, Gupta MM (2008). Separation and quantification of lignans in *Phyllanthus* species by a simple chiral densitometric method. Journal of Separation Science 31, 23-38.

Tahir I, Khan MR, Shah NA, Aftab M (2016). Evaluation of phytochemicals, antioxidant activity and amelioration of pulmonary fibrosis with *Phyllanthus emblica* leaves. BMC Complementary and Alternative Medicine 16, 406-411.

Tjandrawinata RR, Susanto LW, Nofiarny D (2017). The use of *Phyllanthus niruri* L. as an immunomodulator for the treatment of infectious diseases in clinical settings. Asian Pacific Journal of Tropical Diseases 7, 132–140.

Tram NCT, Son NT, Thao DT, Cuong NM (2016). Kaempferol and kaempferol glycosides from *Phyllanthus acidus* leaves. Vietnam Journal of Chemistry 54, 790–793.

Tung YT, Huang CZ, Lin JH, Yen GC (2018). Effect of *Phyllanthus emblica* L. fruit on methionine and choline-deficiency dietinduced nonalcoholic steatohepatitis. Journal of Food and Drug Analysis 26, 1245–1252.

Unander DW, Webster GL, Blumberg BS (1995). Usage and bioassays in *Phyllanthus* (Euphorbiaceae) IV: clustering of antiviral uses and other effects. Journal of Ethnopharmacology 45, 1–18. doi: 10.1016/0378-8741(94)01189-7

Unigwe CR, Enibe F, Igwe KK, Igwe IR, Stephen NO, Koleosho SA, Balogun FA, Shobowale OM, Okonkwo CJB (2020). Effects of *Phyllanthus amarus* (Stone-Breaker) Leaf Meal Supplementation on Haematology and Serum Biochemistry of Broiler Chickens. Direct Research Journal of Biology and Biotechnology, 6(5), 57-63. DOI:https//doi.org/10.26765/DRJBB2072442 8

Unigwe CR, Enibe F, Egwu UL, Igwe RI, Shobowale MO, Njoku CP (2021). Effects of *Phyllanthus amarus* on faecal loads of *Salmonella* Enteritidis and castor-oil induced diarrhoea in broiler chickens. Animal Research International 18(2), 4083 – 4093.

Uzor BC, Umeh LA, Manu OU (2016). Phytochemical Composition and Antimicrobial Potential of *Phyllanthus amarus* Leaf Extract Against Some Clinical Isolates. Nigerian Journal of Microbiology 30(2), 3464-3467

Wagle N, Nagarjuna S, Sharma H, Dangi NB, Sapkota HP, Naik BS, Padhaya RR (2016). Evaluation of antinociceptive and antiinflammatory activity of phytosterol present in chloroform extract of *Phyllanthus maderaspatensis*. Indian Journal of Physiology and Pharmacology 60, 90–95.

Wu Y, Xie SS, Hu ZX, Wu ZD, Guo Y, Zhang JW, Wang JP, Xue YB (2017). Triterpenoids from Whole Plants of *Phyllanthus urinaria*. Chinese Herbal Medicine 9, 193–196.

Yan H, Han LR, Zhang X, Feng JT (2017). Two new Anti-TMV active chalconoid analogues from the root of *Phyllanthus emblica*. Natural Product Research 31, 2143– 2148.

Yang B, Kortesniemi M, Liu P, Karonen M, Salminen JP (2012). Analysis of hydrolyzable tannins and other phenolic compounds in emblic leafflower (*Phyllanthus emblica* L.) fruits by high performance liquid chromatography–electrospray ionization mass

JCBR Vol 3 Is 4 July-Aug 2023

spectrometry. Journal of Agricultural and Food Chemistry 60, 8672–8683.

Yoon WH, Lee KH (2017). Antiinflammatory, Anti-arthritic and Analgesic Effect of the Herbal Extract Made from *Bacopa monnieriis, Cassia fistula* and *Phyllanthus polyphyllus*. Natural Product Science 23, 108–112.

Zhang X, Xia Q, Yang G, Zhu D, Shao Y, Zhang J, Cui Y, Wang R, Zhang L (2017). The anti-HIV-1 activity of polyphenols from *Phyllanthus urinaria* and the pharmacokinetics and tissue distribution of its marker compound, gallic acid. Journal of Traditional Chinese Medical Sciences 4, 158– 166.

Zhang Y, Zhao L, Guo X, Li C, Li H, Lou H, Ren D (2014). Chemical constituents from *Phyllanthus emblica* and the cytoprotective effects on H2O2-induced PC12 cell injuries. Archives of Pharmacal Research 39, 1202– 1211.